

## CALCIUM CHANNELS AND THEIR INVOLVEMENT IN CARDIOVASCULAR DISEASE

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**Abstract**—The widespread distribution of L-type  $\text{Ca}^{2+}$  channels in the cardiovascular system makes that system a natural 'target' for drugs which inhibit L-type  $\text{Ca}^{2+}$  channel activity. Now that tissue-dependent differences in the chemical composition of the calcium antagonist binding sites have been recognized it may be possible to develop drugs with enhanced tissue selectivity. The search for new compounds should not be restricted to improvements in tissue selectivity, however. Some of the ancillary properties of the L-type  $\text{Ca}^{2+}$  channel inhibitors—including their ability to protect against lipid peroxidation—should not be lost because these ancillary properties may contribute significantly to their usefulness as therapeutic agents.

Ion-conducting channels which normally admit extracellular  $\text{Ca}^{2+}$  and which are voltage sensitive and relatively selective for  $\text{Ca}^{2+}$  are present in most cells [1]. Based on their pharmacological and physiological profiles it has been customary to subclassify these channels into L, T or N subtypes [2] but with the benefit of hindsight this classification may be too restrictive. For example, it is now known that dendrites of cerebellar Purkinje cells contain voltage-sensitive,  $\text{Ca}^{2+}$ -selective channels, the pharmacological and biophysical properties of which prevent them from being classified in this way. Currently these  $\text{Ca}^{2+}$  channels are known as P-type channels [3] and one of their distinguishing peculiarities is that, unlike the classical L, T, or N type  $\text{Ca}^{2+}$  channels, they are sensitive to the toxin of the funnel-web spider [4]. There are probably other types of  $\text{Ca}^{2+}$  channel but this paper is concerned primarily with the L-type  $\text{Ca}^{2+}$  channel, since it is this type of  $\text{Ca}^{2+}$  channel which abounds in the cardiovascular system.

L-type  $\text{Ca}^{2+}$  channels are easily identified in terms of their relatively large capacitance, high threshold of activation and sensitivity to a wide variety of organic and inorganic antagonists, including (Table 1) the recently identified polypeptide calciseptine [5]. This particular polypeptide contains 60 amino acids, eight of which are cysteine residues (Table 2). It is present in the venom of the black mamba snake and is the only known naturally occurring polypeptide which specifically antagonizes L-type  $\text{Ca}^{2+}$  channel activity.

Within the last few years the L-type  $\text{Ca}^{2+}$  channel has been isolated and cloned. Its subunits have been identified and its amino acid composition—including some tissue-dependent peculiarities—determined [6]. In addition abnormalities in  $\text{Ca}^{2+}$  channel structure and function have been identified as being associated with certain naturally occurring and drug-induced pathological conditions [7].

### *The structure of the $\text{Ca}^{2+}$ antagonist L-type $\text{Ca}^{2+}$ channel*

The mammalian voltage-dependent, calcium antagonist-sensitive  $\text{Ca}^{2+}$  channel is an oligomeric structure consisting of five putative subunits—designated  $\alpha_1$ ,  $\alpha_2$ ,  $\beta$ ,  $\gamma$  and  $\delta$  [8]. Figure 1 illustrates its probable method of assembly [8]. Of these subunits it is the  $\alpha_1$  subunit which almost certainly constitutes the ion-conducting component of the complex because, when incorporated into planar lipid bilayers [8–11] or microinjected into cells which lack an endogenous  $\alpha_1$  subunit [12], it functions as a calcium-antagonist sensitive,  $\text{Ca}^{2+}$ -conducting channel. This  $\alpha_1$  component of the channel also contains the voltage sensor for the channel [13].

The  $\alpha_1$  subunit of the channel is not extensively glycosylated and it is strongly hydrophobic. Its precise amino acid composition shows some tissue specificity. For example, in cardiac and aortic smooth muscle cells there is a close similarity in composition (93% homology, Table 3), indicating that the aortic smooth muscle  $\alpha_1$  subunit closely resembles its cardiac muscle equivalent (Table 3). However, the aortic, cardiac and brain  $\alpha_1$  subunits are only between 65 and 75% homologous with their skeletal muscle equivalents [6, 14].

Irrespective of whether the  $\alpha_1$  subunit belongs to a  $\text{Ca}^{2+}$  channel complex associated with cardiac, vascular smooth muscle or even skeletal muscle, the amino acids which constitute the subunit are arranged in four repeating hydrophobic motifs each of which contains six membrane spanning segments (Fig. 2). One of the most remarkable features of this arrangement—apart from its homology with the  $\text{Na}^+$  channel [15]—is that segment S4 of each motif appears to be highly conserved in terms of its amino acid composition, in particular with respect to the presence of an arginine or lysine residue at the third position.

It is beyond the scope of this paper to discuss either the biochemistry or the functional significance of the other subunits ( $\alpha_2$ ,  $\gamma$ ,  $\beta$ , and  $\delta$ ) of the  $\text{Ca}^{2+}$

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Table 1. Characteristics of voltage-dependent L-, T- and N-type calcium channels

| Property   | Channel Type |             |             |
|--|--------------|-------------|-------------|
|  | L            | T           | N           |
| Conductance (ps)   | 25           | 8           | 12–20       |
| Activation threshold   | High         | Low         | High        |
| Inactivation rate  | Slow         | Fast        | Moderate    |
| Sensitivity to calcium antagonists:<br>phenylalkylamines,<br>benzothiazepines,<br>dihydropyridines |              |             |             |
| Sensitivity to calciseptine*   | Sensitive    | Insensitive | Insensitive |

\* Calciseptine, a polypeptide, is a naturally occurring calcium antagonist (see Table 2).

Table 2. Chemical composition of calciseptine

| Amino Acid               | Key | Amino Acid              | Key |
|--------------------------|-----|-------------------------|-----|
| Arginine                 | R   | Arginine                | R   |
| Isoleucine               | I   | Glutamic acid           | E   |
| Cysteine                 | C   | Tyrosine                | Y   |
| Tyrosine                 | Y   | Isoleucine              | I   |
| Isoleucine               | I   | Serine                  | S   |
| Histidine                | H   | Glutamic acid           | E   |
| Lysine                   | K   | Arginine                | R   |
| Alanine                  | A   | Glycine                 | G   |
| Serine                   | S   | Cysteine                | C   |
| <sup>10</sup> Leucine    | L   | <sup>40</sup> Glycine   | G   |
| Proline                  | P   | Cysteine                | C   |
| Arginine                 | R   | Proline                 | P   |
| Alanine                  | A   | Threonine               | T   |
| Threonine                | T   | Alanine                 | A   |
| Lysine                   | K   | Methionine              | M   |
| Threonine                | T   | Tryptophan              | W   |
| Cysteine                 | C   | Proline                 | P   |
| Valine                   | V   | Tyrosine                | Y   |
| Glutamic acid            | E   | Glutamine               | Q   |
| <sup>20</sup> Asparagine | N   | <sup>50</sup> Threonine | T   |
| Threonine                | T   | Glutamic acid           | E   |
| Cysteine                 | C   | Cysteine                | C   |
| Tyrosine                 | Y   | Cysteine                | C   |
| Lysine                   | K   | Lysine                  | K   |
| Methionine               | M   | Glycine                 | G   |
| Phenylalanine            | F   | Aspartic acid           | D   |
| Isoleucine               | I   | Arginine                | R   |
| Arginine                 | R   | Cysteine                | C   |
| Threonine                | T   | Asparagine              | N   |
| <sup>30</sup> Glutamine  | Q   | <sup>60</sup> Lysine    | K   |

Table 3. Tissue specificity of the  $\alpha_1$  subunit

| Origin               | % Homology relative to |                     |
|----------------------|------------------------|---------------------|
|                      | Cardiac $\alpha_1$     | Skeletal $\alpha_1$ |
| Cardiac muscle       | —                      | 65                  |
| Aortic smooth muscle | 93                     | 66                  |
| Brain                | —                      | 75                  |
| Skeletal             | 65                     | —                   |

- mice born with the genetically determined and fatal condition of muscular dysgenesis lack functional  $\alpha_1$  receptors in their skeletal but not cardiac  $\text{Ca}^{2+}$  channel complexes [16] which is indicative of different gene coding for the two tissues.
- mRNA blot analysis with probes for the skeletal muscle  $\alpha_1$  subunit show the expected strong band for skeletal muscle, only a weak hybridizing signal for cardiac muscle DNA and no signal for ileal or brain tissue [17].
- Injecting cDNA plasmids coding for cardiac  $\alpha_1$  subunits into skeletal muscle myotubes obtained from dysgenic mice which lack functional  $\alpha_1$  subunits results in the restoration of  $\alpha_1$  subunit activity as indicated by the restoration of  $\text{Ca}^{2+}$  current activity but the  $\text{Ca}^{2+}$  currents are now of the cardiac type, and the tissue is dependent on the presence of extracellular  $\text{Ca}^{2+}$  for excitation-contraction coupling [16].
- The amino acid composition of the terminal intracellular carboxy tail (Fig. 2) of the aortic  $\alpha_1$  subunit is different from that obtained for cardiac (or skeletal) muscle [6].

channel complex, other than pointing out that they probably provide sites for glycosylation and cyclic AMP- and GMP-dependent regulation, as well as stabilizing the  $\text{Ca}^{2+}$  channel complex in the membrane. Of relevance to the present paper, however, is the tissue specificity of the  $\alpha_1$  subunit.

#### *Tissue specificity of the $\alpha_1$ subunit*

Arguments favouring the hypothesis of tissue specificity with respect to this subunit include the following:

In general, therefore, although the  $\alpha_1$  subunit of the  $\text{Ca}^{2+}$  channel complex is well preserved in brain, cardiac, vascular smooth and skeletal muscle; there are subtle differences in the subunit between the different muscle types, including differences in amino acid composition and the length of the carboxy terminus [6]. These differences may contribute to the tissue specificity which some of the prototype calcium antagonists display because it is the  $\alpha_1$  subunit which selectively binds these drugs [18].

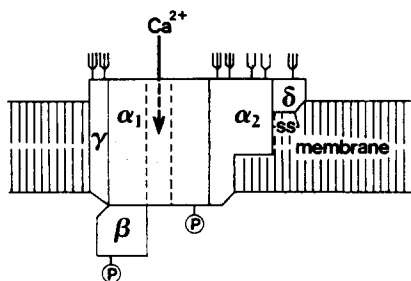


Fig. 1. Schematic representation of the subunit structure of the calcium channel complex. Note that the  $\alpha_1$  subunit contains binding sites for the dihydropyridine-, phenylalkylamine- and benzothiazepine-based calcium antagonists [8].

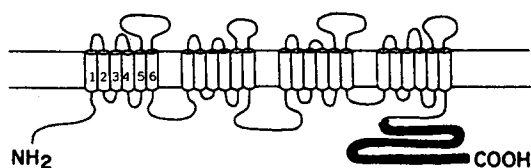


Fig. 2. Schematic representation of the organization of the DHP receptor as it occurs in cardiac and skeletal muscle. The heavily shaded area denotes the carboxy terminus. Note that there are four homologous units, each of which contains six putative transmembrane units.

#### *The characteristics of the calcium antagonist binding sites*

Whilst it is well accepted that the  $\alpha_1$  subunit contains specific high affinity binding sites for the calcium antagonists, the precise location of these sites within the architecture of the subunit is less certain. However, based on the presence or absence of 'use' dependency, it is reasonable to argue that drugs which require the channel to be in the 'open' state and which therefore exhibit 'use' dependency must approach their binding sites by way of an open channel lumen. By contrast, drugs which lack 'use'-dependency or display it to only a small degree probably approach their receptors mainly by way of the lipid-containing membranes, and therefore are usually more lipophilic. The dihydropyridines generally belong to the latter group—with the exception of amlodipine which, presumably because of its charge [19], is slow to penetrate the membrane and hence only slowly reaches equilibrium binding [20]. This may provide this particular calcium antagonist with an advantage because its slow onset of action allows sufficient time for resetting of the baroreceptors, thereby avoiding some of the reflex-induced changes in heart rate which are associated with other dihydropyridine-based antagonists and which often require ancillary treatment in the form of  $\beta$ -adrenoceptor antagonist therapy.

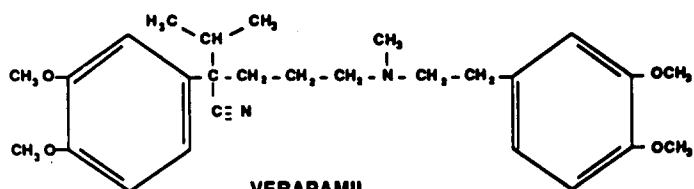
Binding studies using tissue homogenates from heart, smooth muscle, skeletal muscle and brain have clearly established that the 'classical' calcium

antagonists—the dihydropyridines, phenylalkylamines and benzothiazepines—occupy distinct but allosterically coupled binding sites on the channel protein [18, 21]. There may, however, be many more binding sites than have been recognized to date. These other sites need not necessarily be associated with the  $\alpha_1$  subunit. Alternatively, they may be located on hitherto unrecognized sites of the subunit. One of the major reasons for arguing that other binding sites may exist is that new ligands are appearing which are chemically unrelated to the prototypes, verapamil, nifedipine and diltiazem (Fig. 3). Although these new ligands show all the pharmacological and physiological characteristics of prototype calcium channel antagonists, they appear not to bind to recognized binding sites in the  $\alpha_1$  complex. Examples of these novel compounds include the compounds pimozide and HOE 166 (Fig. 4).

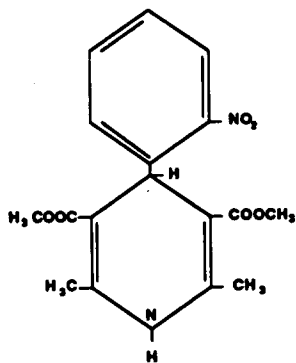
#### *Novel calcium antagonists with additional antagonist activity*

A relatively new development in the field of calcium antagonism, and one which is relevant to the cardiovascular system, is the deliberate search for compounds which can be used to achieve the simultaneous antagonism of other receptors in addition to that of the voltage-sensitive  $\text{Ca}^{2+}$  channels but within the same dose range as these channels. Two examples serve to illustrate this point: AHR-16303B, a joint calcium channel and 5-HT<sub>2</sub> receptor antagonist [22]; and naftopidil, a joint  $\alpha_1$  adrenoceptor and calcium antagonist, being developed particularly for vascular smooth muscle [23]. The search for new calcium antagonists with ancillary properties may be indicative of the trends in this field.

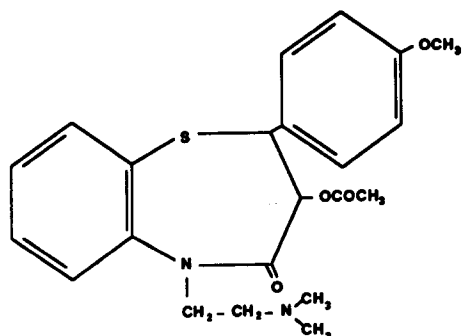
Before discussing the specific topic of this paper—the involvement of calcium channels in cardiovascular disease as indicated by calcium antagonist therapy—it may be useful to summarize the main trends which are being followed in the search for new compounds. One such trend has already been mentioned, i.e. the search for potent calcium antagonists with ancillary antagonist properties, for example, the combined 5-HT<sub>2</sub> receptor and calcium channel antagonist, AHR-16303B, mentioned above [22]. Other trends include the search for tissue-selective drugs and the development of more potent, long-acting compounds. Nimodipine, a dihydropyridine developed for use in the management of patients with stroke or subarachnoid haemorrhage provides an example of the former class [24]. Its use under these conditions depends largely on its selectivity for the cerebral vasculature and its ability to cross the blood-brain barrier. Its relative selectivity for the cerebral vasculature ensures that its introduction is not accompanied by a sustained reduction in peripheral vascular resistance—an effect which would exacerbate damage caused by inadequate cerebral perfusion. Its efficacy depends in part upon its cerebral vasodilator activity, which may improve perfusion in the affected area, and its cytoprotective effect [25]. An example of a long-acting drug is provided by the more recently developed



mol wt 454.59



mol wt 346.34



mol wt 414.52

Fig. 3. Chemical structure of the three prototype calcium antagonists verapamil, diltiazem and nifedipine.

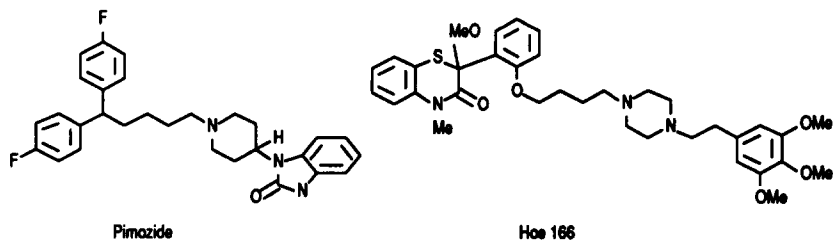


Fig. 4. Chemical structure of two novel calcium antagonists structurally unrelated to the prototype drugs.

dihydropyridine, amlodipine. Amlodipine dissociates only slowly from its receptor [20], a property which facilitates the use of this particular calcium antagonist on a once-a-day basis.

#### *Calcium channels and their involvement in cardiovascular disease*

Calcium ion influx by way of the voltage-activated, calcium antagonist-sensitive channels plays a major role in a wide spectrum of cardiovascular disorders ranging from hypertension and certain other peripheral vascular disorders to myocardial infarction, angina pectoris, congestive heart failure and, although possibly indirectly, atherosclerosis. The involvement of L-type  $\text{Ca}^{2+}$  channel activity in some of these conditions is outlined below.

**Hypertension.** The primary feature of hypertension is a raised systemic vascular resistance [26]. Any change which may occur in cardiac output must, therefore, be relegated to being a consequence of this resistance and not a cause of the hypertensive state.

Calcium ions play a key role in smooth muscle cell contraction, just as they do in that of cardiac and skeletal muscle. As far as vascular smooth muscle is concerned, "activator  $\text{Ca}^{2+}$ " is derived from two sources; one extracellular and the other intracellular. The extracellular  $\text{Ca}^{2+}$  can gain access to the cytosol in several different ways, including by way of the L-type  $\text{Ca}^{2+}$  channels and, but to a lesser extent, the  $\text{Na}^+:\text{Ca}^{2+}$  exchanger. As far as the intracellularly derived  $\text{Ca}^{2+}$  is concerned, release is from the sarcoplasmic reticulum and is in response to either the  $\text{Ca}^{2+}$  which is entering by way of the L-type channels or enhanced inositol triphosphate metabolism, such as that caused by endothelin-1 or  $\alpha_1$  adrenoceptor stimulation. Some vascular smooth muscle cells are more richly endowed with sarcoplasmic reticulum than others. For example, the large conduit vessels contain more reticulum than the small resistance vessels [27]. As a result, the small resistance vessels are more heavily dependent on the influx of  $\text{Ca}^{2+}$  than their larger conduit counterparts. It is these small resistance vessels which are the primary target of calcium antagonists designed for use as blood pressure-lowering agents.

Of the presently available calcium antagonists the dihydropyridine-based drugs exhibit greater selectivity for the vasculature than their phenylalkylamine- or benzothiazepine-based counterparts. Felodipine and nitrendipine, for example, are relatively selective for the vasculature, whereas the phenylalkylamine-based calcium antagonist verapamil lacks this selectivity and is equipotent in producing  $\text{Ca}^{2+}$  channel blockade in the heart and vasculature. This relative selectivity of the dihydropyridines is multifactorial in origin. Factors which are involved include: (a) the chemistry of the compounds; (b) the relatively low transmembrane potential difference ( $-50$  mV for smooth muscle, and  $-85$ – $90$  mV for most cardiac muscle cells [28]); and (c) the unusual amino acid content of the  $\alpha_1$  subunit of the  $\text{Ca}^{2+}$  channel complex, as already described.

**The pathophysiology of hypertension.** The major

causes of hypertension can be divided into at least five categories: (a) the presence of humoral constrictors, some of which (for example, endothelin-1) may directly or indirectly activate L-type  $\text{Ca}^{2+}$  channels; (b) pressure-induced structural changes in the resistance vessel, causing a reduction in lumen diameter; (c) sympathetic overdrive; (d) autotoxins, paracoids and mitogens associated with blood vessels; (e) a relative increase in the number of functioning L-type  $\text{Ca}^{2+}$  channels [29]; and (f) the prolonged activation of these channels [30]. Any hypothesis which is aimed at accounting for the hypertensive state must also take into account the following: (a) endothelium-dependent relaxation is impaired [31]; (b) the smooth muscle cells of the vasculature develop a hypersensitivity towards agonists which evoke a contractile response [32]; and (c) cytosolic  $\text{Ca}^{2+}$  is raised [32]. Under these circumstances, partial inhibition of the L-type  $\text{Ca}^{2+}$  channels should be expected to cause vasodilation, particularly in hypertensives. As an added bonus, some of the calcium antagonists also slow  $\text{Na}^+$  reabsorption in the kidney thereby attenuating any gain in  $\text{Ca}^{2+}$  which might subsequently occur by way of the  $\text{Na}^+:\text{Ca}^{2+}$  exchanger. By far the most attractive hypothesis which is available at the moment, however, and which is supported by experimental data is that the blood pressure-lowering activity of the calcium antagonists in hypertensives is due to their ability to attenuate the prolonged activation of the smooth muscle L-type  $\text{Ca}^{2+}$  channel, which appears to be peculiar to the vascular smooth muscle cell of hypertensives [30].

Hence, as far as hypertension is concerned L-type  $\text{Ca}^{2+}$  channel density is increased [29] and the channels have a prolonged duration of opening [30]. The end result is a raised cytosolic  $\text{Ca}^{2+}$  and hence an increase in peripheral vascular resistance which will be exacerbated by circulating or locally released agents (angiotensin, endothelin-1, vasopressin) which either activate the L-type  $\text{Ca}^{2+}$  channels or exacerbate internal  $\text{Ca}^{2+}$  release.

**Myocardial infarction.** Ischaemia-induced myocardial injury which progresses to an irreversible loss of structure and function (infarction) is the least desirable consequence of inadequate coronary perfusion. The other two conditions, "stunning" [33] and "hibernation" [34], are reversible conditions, and in contrast to infarction do not result in irreversible calcium overload and cell necrosis [35].

$\text{Ca}^{2+}$  influx through the L-type  $\text{Ca}^{2+}$  channels contributes to the onset of ischaemic conditions in several ways. Firstly, these channels provide one of the routes for  $\text{Ca}^{2+}$  entry prior to loss of membrane integrity, and at a time when the energy stores are depleted to such an extent [36] as to limit extrusion of the  $\text{Ca}^{2+}$  across the sarcolemma or its retrieval by the sarcoplasmic reticulum. Under these conditions, cytosolic  $\text{Ca}^{2+}$  rises (Fig. 5) with the resultant activation of  $\text{Ca}^{2+}$ -dependent protein kinases, phospholipases and proteases [37]. Secondly, it is the  $\text{Ca}^{2+}$  which enters by way of the L-type  $\text{Ca}^{2+}$  channels which dictates largely how rapidly the high energy phosphate reserves become depleted, since it is the  $\text{Ca}^{2+}$ -induced activation of energy which is the main determinant of myosin ATPase usage under

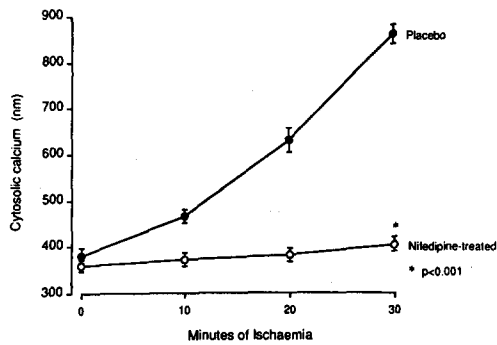


Fig. 5. Cytosolic  $\text{Ca}^{2+}$  (measured by NMR spectroscopy using F-BAPTA as a  $\text{Ca}^{2+}$  indicator) in isolated rat hearts made ischaemic for 30 min. Note that in the nifedipine series the drug was added to the perfusion buffer 15 min before the hearts were made ischaemic [46].

these conditions and early during the ischaemic event, when active tension development is still taking place, this is the major source of energy usage.

If  $\text{Ca}^{2+}$  entry by way of the L-type  $\text{Ca}^{2+}$  channels plays an important role in the sequence of events which results in infarction and if ionic homeostasis with respect to  $\text{Ca}^{2+}$  is lost early during the ischaemic episode (Fig. 5), then it is logical to assume that calcium antagonist therapy will be useful only if the drugs are used prophylactically. Two large scale clinical trials [38, 39] have confirmed this assumption.

At this stage, and before considering the criteria which are applicable to the choice of calcium antagonists most suitable for use in the management of patients with ischaemic heart disease, it may be useful to digress and consider the possible involvement of  $\text{Ca}^{2+}$  channels which are not of the L-type.

Recent evidence [40] indicates that ischaemic episodes may have a profound effect on the  $\text{Ca}^{2+}$ -storing capacity of the sarcoplasmic reticulum. Not only is this organelle 'attacked' by the oxy- and hydroxy-radicals which are generated during prolonged episodes of ischaemia, and subsequent reperfusion [41], the functioning of its  $\text{Ca}^{2+}$  release channels is also affected [42]. Probably because of ATP depletion, these 'release' channels exhibit a prolonged opening time thereby facilitating efflux into the cytosol of  $\text{Ca}^{2+}$  which this organelle had already retrieved from the cytosol. Hence, at a time when the sarcolemmal L-type  $\text{Ca}^{2+}$  channels are still functioning, although their density may be reduced [43], intracellular homeostasis with respect to  $\text{Ca}^{2+}$  is lost, partly because of insufficient ATP to fuel the  $\text{Ca}^{2+}$  ATPases responsible for expelling  $\text{Ca}^{2+}$  from the cytosol and, secondly, because of the failure of the  $\text{Ca}^{2+}$  release channels of the reticulum to remain closed. Not surprisingly, cytosolic  $\text{Ca}^{2+}$  rises (Fig. 5) but only in part because of  $\text{Ca}^{2+}$  entry by way of the L-type  $\text{Ca}^{2+}$  channels.

Before considering the involvement of the L-type  $\text{Ca}^{2+}$  channels in other cardiovascular disorders it may be useful to consider briefly the properties a calcium antagonist should possess if it is to be

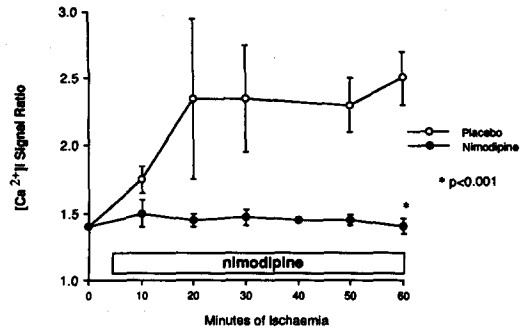


Fig. 6. Effect of nimodipine given as an infusion starting 10 min before occluding the middle cerebral artery in cats (data from Ref. 44; diagram from Ref. 47). Nimodipine was infused at a rate of  $1 \mu\text{g}/\text{kg}/\text{min}$ . Indo 1-AM was used as the  $\text{Ca}^{2+}$ -indicator.

developed for use in the management of patients with ischaemic heart disease. Probably, the drug should: (a) lack marked negative inotropy. Otherwise the pumping capacity of the affected myocardium may be further reduced, with the danger of precipitating cardiac failure. (b) It should show some preference for the coronary vasculature. (c) It should reduce peripheral vascular resistance (to reduce the workload on the heart) but not cause such profound vasodilation as to either trigger a reflex tachycardia or further reduce blood flow through coronary vessels which still remain patent. (d) It should have a long half-life. This is achieved by the use of slow release formulations or, as in the case of amlodipine [20], is due to a slow rate of dissociation from its receptor.

#### Cerebral ischaemia

Cerebral ischaemia can occur for a variety of reasons including a precipitous fall in blood pressure, emboli arising from thrombi or an ulcerated or ruptured plaque, sustained vasospasm, and cerebral haemorrhage. The term 'ischaemic stroke' is often used to describe the acute event in which damage to the brain is immediate. In this respect, the condition is quite different from that caused by a subarachnoid haemorrhage where the damage to the brain is delayed and most probably due to secondary vasospasm [44]. Irrespective of which of these conditions prevails, oxyradicals accumulate and cytosolic  $\text{Ca}^{2+}$  rises [45] with  $\text{Ca}^{2+}$  overload, cell death and tissue necrosis the inevitable outcome. Calcium ion flux through  $\text{Ca}^{2+}$ -selective channels must contribute to this scenario because nimodipine [45], the  $\text{Ca}^{2+}$  antagonist which shows some selectivity for the cerebral vasculature and which penetrates across the blood-brain barrier, attenuates the rise in cell  $\text{Ca}^{2+}$  caused by an occlusive event (Fig. 6).

#### Calcium channels and the atherogenic process

Atherosclerosis is a multifactorial disease which involves not just lipid accumulation but also the localized deposition of collagen, elastin and calcium [48, 49]. The proposed use of calcium antagonists as

anti-atherogenic agents was based initially on the assumption that the  $\text{Ca}^{2+}$  which accumulates within the mature plaque involves  $\text{Ca}^{2+}$  entry by way of the L-type channels in the vascular smooth muscle cells [50]. Such an assumption may be correct, but  $\text{Ca}^{2+}$ -sensitive processes are involved in many of the events of the early stages of lesion formation including monocyte adhesion, release of growth factors, smooth muscle cell proliferation and platelet aggregation. Inhibitors of L-type  $\text{Ca}^{2+}$  channels are now known to have a direct effect on many of the events which are involved in the formation of atherogenic lesions. For example, they inhibit the migration [51] and proliferation [52] of vascular smooth muscle cells; collagen and matrix synthesis [53]; and cholesteryl ester deposition [54]; and protect against lipid peroxidation-induced injury [55], an event which may play a pivotal role in the uptake of low density lipoprotein during the initial stages of lesion formation [55, 56]. Hence, although inhibitors of L-type  $\text{Ca}^{2+}$  channels are anti-atherogenic [49] it does not necessarily follow that L-type  $\text{Ca}^{2+}$  channels are involved in any major way in the early stages of lesion formation, because the presently available inhibitors of L-type  $\text{Ca}^{2+}$  channels have ancillary properties, some of which may be of therapeutic importance.

#### The coronary circulation

L-type  $\text{Ca}^{2+}$  channels are present in the coronary vasculature in both the large conduit vessels and the small resistance arteries [50]. Activation of these channels, irrespective of whether it be direct or the indirect consequence of locally released endothelin [57], probably does contribute to or is the major cause of coronary vasospasm.

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#### REFERENCES

1. Bean BP, Classes of calcium channels in vertebrate cells. *Annu Rev Physiol* 51: 367–384, 1989.
2. Miller R, Multiple calcium channels and neuronal function. *Science* 235: 46–52, 1987.
3. Llinas R, Sugimori M, Lin J-W and Cherksey B, Blocking and isolation of a calcium channel from neurons in mammals and cephalopods utilizing a toxin fraction (FTX) from funnel web spider poison. *Proc Natl Acad Sci USA* 86: 1689–1693, 1989.
4. Lin J-W, Rudy B and Llinas R, Funnel-web spider venom and a toxic fraction block calcium current expressed from rat brain mRNA in *Xenopus* oocytes. *Proc Natl Acad Sci USA* 87: 4538–4542, 1990.
5. de Weille JR, Schweitz H, Maes P, Tartar A and Lazdunski M, Calciseptine, a peptide isolated from black mamba venom, is a specific blocker of the L-type calcium channel. *Proc Natl Acad Sci USA* 88: 2437–2440, 1991.
6. Koch WJ, Hui A, Shull G, Ellinor P and Schwartz A, Characterization of cDNA clones encoding two putative isoforms of the  $\alpha_1$  subunit of the dihydropyridine-sensitive calcium channel isolated from rat brain and rat aorta. *FEBS Lett* 250: 386–388, 1989.
7. Nayler WG (Ed.), "Up" and "down" regulation of the calcium antagonist binding sites. In: *The Second*

*Generation of Calcium Antagonists*, pp. 55–66. Springer, Heidelberg, 1991.

8. Catterall WA, Seagar MJ, Takahashi M and Nunoki K, Molecular properties of dihydropyridine-sensitive calcium channels. *Ann NY Acad Sci* 560: 1–14, 1989.
9. Flockerzi V, Oeken HJ, Hofmann F, Pelzer D, Cavalié A and Trautwein W, Purified dihydropyridine-binding site from skeletal muscle T-tubules is a functional calcium channel. *Nature* 323: 66–68, 1986.
10. Curtis BM and Catterall WA, Reconstitution of the voltage-sensitive calcium channel purified from skeletal muscle transverse tubules. *Biochemistry* 25: 3077–3083, 1986.
11. Smith JS, Rousseau E and Meissner G, Calmodulin modulation of single sarcoplasmic reticulum  $\text{Ca}^{2+}$ -release channels from cardiac and skeletal muscle. *Circ Res* 64: 352–359, 1989.
12. Knudson CM, Imagawa T, Kahl SD, Gauer MG, Leung AT, Sharp AH, Jay SD and Campbell KP, Evidence for physical association between junctional sarcoplasmic reticulum ryanodine receptor and junctional transverse tubular dihydropyridine receptor. *Biophys J* 53: 605a, 1988.
13. Tanabe T, Beam KG, Adams BA, Niidome T and Numa S, Regions of the skeletal muscle dihydropyridine receptor critical for excitation-contraction coupling. *Nature* 346: 567–569, 1990.
14. Koch WJ, Ellinor PT and Schwartz A, cDNA cloning of a dihydropyridine sensitive calcium channel from rat aorta. *J Biol Chem* 265: 17786–17791, 1990.
15. Noda M, Ikeda T, Kayano T, Suzuki H, Takeshima H, Kurasaki M, Takahashi H and Numa S, Existence of distinct sodium channel messenger RNA's in rat brain. *Nature* 320: 118–192, 1986.
16. Mikami A, Imoto K, Tanabe T, Niidome T, Mori Y, Takeshima H, Narumiya S and Numa S, Primary structure and functional expression of the cardiac dihydropyridine-sensitive calcium channel. *Nature* 340: 230–233, 1989.
17. Ellis SB, Williams ME, Ways NR, Brenner R, Sharp AH, Leung AT, Campbell KP, McKenna E, Koch WJ, Hui A, Schwartz A and Harpold MM, Sequence and expression of mRNA's encoding the  $\alpha_1$  and  $\alpha_2$  subunits of a DHP-sensitive calcium channel. *Science* 241: 1661–1664, 1988.
18. Glossman H and Streissnig J, Molecular properties of calcium channels. *Rev Physiol Biochem Pharmacol* 114: 1–105, 1990.
19. Burgess RA, Carter AJ, Gardiner DG and Higgins AJ, Amlodipine, a new dihydropyridine calcium channel blocker with slow onset and duration of action. *Br J Pharmacol* 82: 281P, 1985.
20. Nayler WG and Gu XH,  $(-)[^3\text{H}]$ Amlodipine binding to rat cardiac membranes. *J Cardiovasc Pharmacol* 17: 587–592, 1991.
21. Godfraind T, Miller R and Wibo M, Calcium antagonism and calcium entry blockade. *Pharmacol Rev* 38: 321–416, 1986.
22. Barrett RJ, Appell KC, Kilpatrick BF, Proakis AG, Nolan JC and Walsh DA, AHR-16303B, a novel antagonist of 5-HT<sub>2</sub> receptors and voltage-sensitive calcium channels. *J Cardiovasc Pharmacol* 17: 41–53, 1991.
23. Himmel HM, Glossmann H and Ravens U, Naftopidil, a new  $\alpha$  adrenoceptor blocking agent with calcium antagonistic properties: characterization of  $\text{Ca}^{2+}$  antagonist effects. *J Cardiovasc Pharmacol* 17: 213–221, 1991.
24. Towart R and Kazda S, The cellular mechanism of action of nimodipine (Baye9736) a new calcium antagonist. *Br J Pharmacol* 67: 409–410, 1979.
25. Scriabine A, Schuurman T and Traber J, Pharmacological basis for the use of nimodipine in central

- nervous system disorders. *FASEB J* 3: 1799–1806, 1989.
26. Page IH, *Hypertensive Mechanisms*. Grune Stratton, Orlando, 1987.
  27. Somlyo AV, Excitation–contraction coupling and the ultrastructure of smooth muscle. *Circulation* 57: 497–507, 1985.
  28. Hirst GDS and Edwards FR, Sympathetic neuroeffector transmission in arteries and arterioles. *Physiol Rev* 69: 546–604, 1989.
  29. Rusch NJ and Hermsmeyer K, Calcium currents are altered in the vascular muscle cell membranes of spontaneously hypertensive rats. *Circ Res* 63: 997–1002, 1988.
  30. Godfraind T, Kazda S and Wibo M, Effects of a chronic treatment of nisoldipine, a calcium antagonistic dihydropyridine, on arteries of spontaneously hypertensive rats. *Circ Res* 68: 674–682, 1991.
  31. Panza JA, Quyyumi AA, Brush JE and Epstein SE, Abnormal endothelium-dependent vascular relaxation in patients with essential hypertension. *N Eng J Med* 323: 22–27, 1990.
  32. Pagageorgiou P and Morgan KG, Increased  $\text{Ca}^{2+}$  signalling after  $\alpha$  adrenoceptor activation in vascular hypertrophy. *Circ Res* 68: 1080–1084, 1991.
  33. Braunwald E and Kloner RA, The stunned myocardium: prolonged, postischemic ventricular dysfunction. *Circulation* 66: 1146–1149, 1982.
  34. Rahimtoola SH, The hibernating myocardium. *Am Heart J* 117: 211–221, 1989.
  35. Shen AC and Jennings RB, Kinetics of calcium accumulation in acute myocardial ischemic injury. *Am J Pathol* 67: 441–452, 1972.
  36. Jennings RB and Reimer KA, The cell biology of acute myocardial ischemia. *Annu Rev Med* 42: 225–246, 1991.
  37. Nayler WG, Panagiotopolous S, Elz JS and Sturrock WJ, Fundamental mechanisms of action of calcium antagonists in myocardial ischemia. *Am J Cardiol* 59: 75B–83B, 1987.
  38. The Danish Study Group on Verapamil in Myocardial Infarction, Effect of verapamil on mortality and major events after acute myocardial infarction (The Danish verapamil infarction trial 11). *Am J Cardiol* 66: 779–785, 1990.
  39. The Multicenter Diltiazem Postinfarction Trial Research Group, The effect of diltiazem on mortality and reinfarction after myocardial infarction. *N Eng J Med* 319: 385–392, 1988.
  40. Hammond B and Hess ML, The oxygen free-radical system: potential mediator of myocardial injury. *J Am Coll Cardiol* 6: 215–220, 1985.
  41. Bolli R, Mechanism of myocardial “stunning”. *Circulation* 82: 723–738, 1990.
  42. Homberg SRM and Williams AJ, Single channel recordings from human cardiac sarcoplasmic reticulum. *Circ Res* 65: 1445–1449, 1989.
  43. Dillon JS and Nayler WG, [ $^3\text{H}$ ]Verapamil binding to rat cardiac sarcolemmal membrane fragments: an effect of ischaemia. *Br J Pharmacol* 90: 99–109, 1987.
  44. Tettenborn D and Dycka J, Prevention and treatment of delayed ischemic dysfunction in patients with aneurysmal subarachnoid hemorrhage. *Stroke* 21 (Suppl IV): IV-85–IV-89, 1990.
  45. Greenberg JH, Vematsu D, Araki N, Hickey WF and Reivich M, Cytosolic free calcium during focal cerebral ischemia and the effects of nimodipine on calcium and histological damage. *Stroke* 21 (Suppl IV): IV-72–IV-77, 1990.
  46. Nayler WG (Ed.), Second generation calcium antagonists and the ischemic myocardium. In: *The Second Generation of Calcium Antagonists*, pp. 129–135. Springer, Heidelberg, 1991.
  47. Nayler WG (Ed.), Calcium antagonists and the management of cerebral ischaemia. In: *The Second Generation of Calcium Antagonists*, pp. 199–208. Springer, Heidelberg, 1991.
  48. Nayler WG (Ed.), Calcium antagonists and atherosclerosis. In: *Calcium Antagonists*, pp. 235–248. Academic Press, London, 1988.
  49. Henry PD, Atherogenesis, calcium and calcium antagonists. *Am J Cardiol* 66: 31–71, 1990.
  50. Fleckenstein A, History of calcium antagonists. *Circ Res* 52 (Suppl 1): I3–I16, 1983.
  51. Nakao I, Ito H, Ooyama T, Chang WC and Murota S, Calcium dependency of aortic smooth muscle cell migration induced by 12-L-hydroxy-5,8,10,14-eicosatetraenoic acid. *Atherosclerosis* 46: 309–319, 1983.
  52. Nilsson I, Sjolund M, Palmberg L, van Euler AM, Jonzon B and Thyborg I, The calcium antagonist nifedipine inhibits arterial smooth muscle cell proliferation. *Atherosclerosis* 58: 109–112, 1985.
  53. Orekhov AN, Tertov VU, Khashimov KA, Kudryashou SA and Smirnov VN, Antiatherosclerotic effects of verapamil in primary culture of human aortic intimal cells. *J Hypertens* 4 (Suppl 6): S153–S155, 1986.
  54. Daugherty A, Rateri DL, Schonfeld G and Sobel BE, Inhibition of cholesteryl ester deposition in macrophages by calcium entry blockers: an effect dissociable from calcium entry blockade. *Br J Pharmacol* 91: 113–118, 1987.
  55. Janero DR and Burghardt B, Antiperoxidant effects of dihydropyridine calcium antagonists. *Biochem Pharmacol* 38: 4344–4348, 1989.
  56. Ondrias K, Misik V, Gergel D and Stasko A, Lipid peroxidation of phosphatidylcholine liposomes depressed by the calcium channel blockers nifedipine and verapamil and by the antiarrhythmic–antihypoxic drug stobadine. *Biochim Biophys Acta* 1330: 238–245, 1989.
  57. Luscher TF, Yang Z, Tschudi M, von Segesser L, Stulz P, Boulanger C, Siebenmann R, Turina M and Buhler FR, Interaction between endothelin-1 and endothelin-derived relaxing factor in human arteries and veins. *Circ Res* 66: 1088–1094, 1990.